

# Phage Display System--Super Electrocompetent Cells

## Product description

**ER2738 Super Electrocompetent Cells** deliver  $\geq 2 \times 10^{10}$  cfu/ug of DNA and are particularly useful for phage display protein expression. ER2738 cells are also suitable for M13 phage work, general cloning, blue/ white screening and protein expression.

**SS320 (MC1061 F') Super Electrocompetent Cells** deliver  $\geq 4 \times 10^{10}$  cfu/ug of DNA and are particularly useful for phage display protein expression. SS320, also known as MC1061 F' cells, are also suitable for M13 phage work, general cloning, blue/white screening and protein expression.

**TG1 Super Electrocompetent Cells** deliver  $\geq 4 \times 10^{10}$  cfu/ug of DNA and are particularly useful for phage display protein expression. TG1 cells are also suitable for M13 phage work, general cloning, blue/white screening and protein expression.

## Product specifications

Strain	Transformation Efficiency	Genotype
ER2738	$\geq 2 \times 10^{10}$ cfu/ug of pUC DNA	<i>[F'proA<sup>+</sup>B<sup>+</sup>lac<sup>q</sup>Δ(lacZ)M15 zzz::Tn10 (tet<sup>r</sup>)] fhuA2 glnVΔ(lac-proAB) thi-1Δ(hsdS-mcrB)5</i>
SS320 (MC1061F')	$\geq 4 \times 10^{10}$ cfu/ug of pUC DNA	<i>[F'proA<sup>+</sup>B<sup>+</sup>lac<sup>q</sup>lacZΔM15Tn10(tet<sup>r</sup>)]hsdRmcrBaraD139Δ(araABC-leu)7679 ΔlacX74 galUgalK rpsL thi</i>
TG1	$\geq 4 \times 10^{10}$ cfu/ug of pUC DNA	<i>[F'traD36proA<sup>+</sup>B<sup>+</sup>lac<sup>q</sup>ZΔM15]supEthi-1Δ(lac-proAB)Δ(mcrB-hsdSM)5(rK- mK-)</i>

## Product designations and kit components

Cat No	Product	Kit Size	Description	Volume
P021	TG1 Super Electrocompetent Cells	12 reactions (DUOS*)	TG1 Super Electrocompetent Cells	12 (12 x 5 μL)
			Transformation Control pUC19	(1 x 20μL)
P022	ER2738 Super Electrocompetent Cells	12 reactions (DUOS*)	ER2738 Super Electrocompetent Cells	12 (12 x 50μL)
			Transformation Control pUC19	(1 x 20μL)
P023	SS320 (MC1061 F') Super Electrocompetent Cells	12 reactions (DUOS*)	SS320 (MC1061 F') Super Electrocompetent Cells	12 (12 x 50μL)
			Transformation Control pUC19	(1 x 20μL)

\*Phage Display Super Electrocompetent Cells are packaged as DUOS in 50ul aliquots, sufficient for two transformations per tube.

\*\*Supercoiled pUC19 DNA (10pg/μL) is provided as a control for transformation and should be stored at -20 to -80 °C. Use 1 μL (10 pg) for transformation.

## Product designations and kit components

Super Electrocompetent cells require storage at -80 °C

## Preparation for transformation

Transformation is carried out in a 0.1 cm gap cuvette using 25μL of cells. Optimal settings for electroporation are listed in Table 1 below. Typical time constants are 3.5 to 4.5 msec.



Condition	Optimal Setting	Alternate Settings (~ 20-50% lower efficiencies)
<b>Cuvette gap</b>	1.0 mm	1.0 mm
<b>Voltage</b>	1,800 V	1,400-1,600 V
<b>Capacitance</b>	10 $\mu$ F	25 $\mu$ F
<b>Impedance</b>	600 Ohms	200 Ohms
<b>Electroporator</b>	Bio-Rad Micro Pulser #165-2100; Bio-Rad E. coli Pulser #165-2102; Bio-Rad Gene Pulser II #165-2105; BTX ECM630 Electroporation System; Eppendorf Electroporator 2510.	

Table 1. Electroporation conditions for Phage Display Super Electrocompetent Cells

## Transformation protocol for cells

To ensure successful transformation results, the following precautions must be taken:

- Ligation reactions must be heat killed at 70 °C for 15 minutes before transformation. The ligation reaction can be used directly after heat inactivation, without purification of the ligation products.
- DNA samples must be purified and dissolved in water or a buffer with low ionic strength, such as TE. The presence of salt in the electroporation sample leads to arcing at high voltage, resulting in loss of the cells and DNA.
- Microcentrifuge tubes and electroporation cuvettes must be thoroughly pre-chilled on ice before use.
- The cells must be completely thawed on ice before use.

1. Have LB Miller and 17 mm x 100 mm sterile culture tubes readily available at room temperature (one tube for each transformation reaction).
2. Place electroporation cuvettes (0.1 cm gap) and microcentrifuge tubes on ice (one cuvette and one microfuge tube for each transformation reaction).
3. Remove Super Electrocompetent Cells from the -80 °C freezer and place on wet ice until they thaw completely (10-15 minutes).
4. When the cells are thawed, mix them by tapping gently. Aliquot 25 $\mu$ L of cells into the chilled microcentrifuge tubes on ice.
5. If using ligation buffer from any LGC Biosearch Technologies cloning or ligation kit, add 1 $\mu$ L of the heat-denatured ligation reaction to the 25 $\mu$ L of cells on ice. Failure to heat-inactivate the ligation reaction will prevent transformation. Stir briefly with pipette tip; do not pipet up and down to mix, which can introduce air bubbles and warm the cells. Use of more than 2 $\mu$ L of ligation mix may cause electrical arcing during electroporation.

For ligation reactions using other commercial kits, please refer to the manufacturer's instructions.

6. Carefully pipet 25 $\mu$ L of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well. Electroporate according to the conditions recommended above.
7. Within 10 seconds of the pulse, add 975 $\mu$ L of LB Miller to the cuvette and pipet up and down three times to resuspend the cells. Transfer the cells and Recovery Medium to a culture tube.
8. Place the tube in a shaking incubator at 250 rpm for 1 hour at 37 °C.
9. Spread up to 100 $\mu$ L of transformed cells on LB(or other nutrient media) agar plates containing the appropriate antibiotic.
10. Incubate the plates overnight at 37 °C.
11. Transformed clones can be further grown in TB or in any other rich culture medium.

## Media recipes

### LB Lennox agar plates

Per litre:

- 10 g tryptone
- 5 g yeast extract
- 5 g NaCl
- 15 g agar



## Medium for growth of transformants

### LB Miller

Per litre:

10 g tryptone

5 g yeast extract

5 g NaCl

Add all components to deionised water. Adjust pH to 7.0 with NaOH. Autoclave and cool to 55 °C.

### TB

Per litre:

11.8 g tryptone

23.6 g yeast extract

9.4 g dipotassium hydrogen phosphate (anhydrous)

2.2 g potassium dihydrogen phosphate (anhydrous)

0.4% glycerol

Add all components to deionised water. Autoclave and cool to 55 °C.

LB Lennox agar is used to maximise colony size. Cells may be plated on LB or other common media—colonies will be noticeably smaller upon comparison but adequate for the vast majority of intents and purposes.

## Technical support and product guarantee

Alpvhhs is dedicated to the success and satisfaction of our customers. Our products are tested to assure they perform as specified when used according to our recommendations. It is imperative that the reagents supplied by the user are of the highest quality. Please follow the instructions carefully and contact our technical service representatives if additional information is necessary. We encourage you to contact us with your comments regarding the performance of our products in your applications. Thank you.

If you require any further support, please do not hesitate to contact our Technical Support Team: [service@alpvhhs.com](mailto:service@alpvhhs.com).

Product guarantee: Alpvhhs guarantees that this product will perform as specified for one year from the date of shipment. Please avoid using reagents for greater than one year from receipt.